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ENVIRONMENTAL HEALTH LABORATORY BRANCH CALIFORNIA DEPARTMENT OF HEALTH SERVICES

QUALITY MANUAL

Environmental Health Laboratory Branch California Department of Health Services 2151 Berkeley Way Berkeley, California 94704

Doc./Rev. No:

QM/R3

Effective Date:

June 15, 1999

Approval:

C. Peter Flessel, Ph.D., Chief, EHLB

Approval:

Michael G. Volz, Ph.D., Quality Assurance Officer

Date

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- 6. The sample air volume (air samples) or surface area (wipe samples) is required for each sample if airborne or surface area concentration is to be reported.

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All proficiency testing samples are delivered to the Quality Assurance Coordinator for inspection for acceptability prior to assignment and log in.

9.3 Sample Acceptability

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mandatory sample submission/analysis request elements described in Section 9.2, criteria for rejection include:

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- 2. Unlabeled or illegibly labeled samples
- 3. Insufficient sample
- 4. Improper container and/or sampling medium
- 5. Lack of, or improper preservation (chemical or temperature)
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- 7. Lack of, or non-intact custody seals (for specific client project samples only)

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- 4. Date Required
- 5. Project ID
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- 10 Group Leader (Section Assignment)
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USEPA REGION 9 LABORATORY RICHMOND, CALIFORNIA

STANDARD OPERATING PROCEDURE #110 SAMPLE RECEIVING AND LOGIN

Signature & Title

epared by:		Date
Nancy J. Wil	Nancy J. Wilson	
viewed by:		
Ken Hendrix	Ken Hendrix	
proved by:		
Brenda Bette	Brenda Bettencourt	
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Periodic Review:		
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USEPA REGION 9 LABORATORY RICHMOND, CALIFORNIA

STANDARD OPERATING PROCEDURE #110 SAMPLE RECEIVING AND LOGIN

Signature & Title

repared by:		
	Nancy J. Wilson	
viewed by:		
Ken Hendri	Ken Hendrix	
oproved by:		
Brenda Bett	Brenda Bettencourt	
Periodic Review:		
Signature	Title	Date
5.9v.	1,112	
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USEPA REGION 9 LABORATORY RICHMOND, CALIFORNIA

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viewed by:		
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USEPA REGION 9 LABORATORY RICHMOND, CALIFORNIA

STANDARD OPERATING PROCEDURE #110 SAMPLE RECEIVING AND LOGIN

Signature & Title

Prepared by:		
	Nancy J. Wilson	
Reviewed by:		
Ken Hendrix	Ken Hendrix	
Approved by:		
Brenda Bette	Brenda Bettencourt	
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David dia Davidana		
Periodic Review:		
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1.0 PURPOSE

The purpose of this SOP is to describe all activities concerning sample receipt and sample login at the USEPA Region 9 Laboratory. Samples are delivered to the Sample Control Center in Room 503. Samples are inspected, unpacked, logged in, labeled and stored in Room 503 or Room 205. Samples are kept refrigerated pending analysis and disposal. Biota samples are kept frozen in Freezer Unit 21 in Room 503 or in the freezer in Room 205 pending analysis and disposal.

2.0 APPLICABILITY

This SOP applies to the receipt of sample matrices including groundwater, surface water, drinking water, waste water, soil, sediment, biota samples and to samples received from USEPA Programs and their grantees throughout Region 9.

3.0 PROCEDURE

3.1 Notification of Sample Receipt

3.1.1 Analytical Requests and Upcoming Sampling Events Tracking Sheet

The Sample Custodian is notified of upcoming samples receipts in two ways. Analytical requests are made available to the Sample Custodian via the ESAT TDF (Environmental Services Assisstance Team Technical Direction Form) file on the Lotus Notes Workspace. Analytical requests (Appendix A) indicate the site, case number, number of samples, matrices, expected arrival dates of samples and analytical support required for a future sampling event. The Upcoming Sampling Events Tracking Sheet (Appendix B) is sent to the Sample Custodian via Lotus Notes electronic mail from the RSCC (Regional Sample Control Cooridinator) giving a more accurate estimated arrival date, numbers of samples and detailing analytical requests for upcoming sampling events.

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3.1.2 Notification of Shipment of Samples

Notification of sample shipments (Appendix C) are sent to the Sample Custodian, via electronic mail or telephone message by the RSCC. Notification includes the shipment air bill number if samples are shipped using an overnight carrier service or expected arrival time if samples are to be hand delivered.

3.1.3 Sample Receipt/Chain of Custody

Samples from the field are delivered by overnight courier or by the sampler. The sample custodian is immediately notified of the sample shipment by the person accepting the shipment.

- 3.1.3.1 If expected shipment is not received, notify the RSCC. The number is posted by the telephone in Room 503. If the RSCC has no explanation, call the courier service and begin tracking the shipment using the air bill number. The RSCC will contact the sampler if the shipment was to be hand delivered.
- 3.1.3.2 Inspect the sample cooler(s) for damage, and examine the integrity of the custody seals on the lid.
- 3.1.3.4 Open the cooler lid and check that the samples are intact and still cold.
- 3.1.3.5 Sign and date the enclosed Chain of Custody form and attached air bill (Appendix D and E).
- 3.1.3.6 Notify the RSCC if there are any problems with the condition of the samples and document notification on Routing Sheet (See section 3.2.2.9.3).

3.1.4 Unpacking and Inspection

Sample containers are usually received in plastic containment bags and glass containers are generally wrapped in bubble wrap to prevent breakage. Samples that are labeled as high concentration or those suspected of containing elevated concentrations of toxic

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materials (packed in paint cans) are unpacked under the fume hood.

- 3.1.4.1 Record the temperature of the samples on the Chain of Custody. If a temperature blank is provided, insert the thermometer into the blank water and allow it to equilibrate for one minute before taking a reading. If no temperature blank has been provided, use the Cole Parmer, 39650-02 Standard Model, Infrared Thermometer by pointing it at the sample label and pulling the trigger. The temperature will be displayed in the digital display window on the top of the thermometer. Both thermometers are stored in the drawer labeled *Thermometers*, in Room 503.
- 3.1.4.2 'Remove the samples from plastic containment bags and packing material.
- 3.1.4.3 Set the samples on the bench in the order they appear on the Chain of Custody form. Set the samples on K-Dry table top soaker paper to absorb condensation moisture.
- 3.1.4.4 If samples arrive broken, wear thick rubber gloves while handling broken sample containers. Discard the broken glass into the cardboard container in Room 503 that has been designated for broken glass. Pour any aqueous sample remaining in either the sample container or the cooler into the carboy labeled for aqueous waste located under the fume hood in Room 503. See SOP Number 125 for detailed instructions for disposal of hazardous waste. Record the identification numbers of the broken samples on the Chain of Custody and the Sample Routing Sheet (see 3.2.4.13). Immediately notify the RSCC with the identities of the broken samples.
- 3.1.4.5 Measure and record the pH of chemically preserved aqueous samples is to be measured and recorded (VOC samples excepted). Fill a 10 mL disposable beaker with sample and immerse a 0-14 pH indicator strip into the aliquot until there is no further color change on the strip. Match the colors on the strip with the colors on the package for the pH of the sample. If a sample pH is other than that indicated for that particular analysis by EPA-600/4-80-055, "Technical Additions to Methods for Chemical Analysis of Water & Waste", immediately notify the RSCC and the Chemistry Team Leader and note any problem on the Sample Routing Sheet (Appendix F). Dispose of the liquid from the disposable beaker in the carboy labeled for aqueous sample waste stored under the fume hood.

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- 3.1.4.6 Preserve drinking water samples with HNO3 to a pH less than 2 if not already done so by sampler (EPA Methods 200.7, 200.8, 200.9).
 - 3.1.4.6.1 In the pH and Turbidity Run Log, enter the date, time, case number, sample number (ABXXXXX), and the initial pH.
 - 3.1.4.6.2 Using 1:1 Nitric Acid and a pipet, add 3 mL of the acid to each 1 liter sample container and re-check the pH. If necessary, add drops of 1:1 Nitric Acid until a pH of <2 is reached. Record the corrected pH in the Run Log in the column titled pH after preservation. Following acidification, the sample should be held for 16 hours before analysis and the pH re-checked. If the sample(s) pH is >2, add more acid and re-check after an additional 16 hours.
 - 3.1.4.6.2 Measure the turbidity for all drinking water samples submitted to the lab for metals analysis. See SOP 401.
- 3.1.4.8 Compare the information on the sample labels with that on the Chain of Custody (sample ID, sample location, date sampled, time sampled). If any information recorded on the sample label fails to match that on the Chain of Custody, immediately notify the RSCC. Resolution of any problems must be written on or accompany the Sample Routing Sheet.

3.2 Sample Login/LIMS

The following procedures provide basic instruction for entering sample information into LABWORKS Enterprise, software dedicated for assigning laboratory identification numbers, maintaining the samples database, generating Routing Forms and sample labels.

3.2.1 Case Definition

A Case usually refers to samples from one sampling event, and may consist of any number of samples. Cases are defined by number, with the case number being assigned by the RSCC.

3.2.1.1 Determine the case number and analyses requested from the ESAT

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TDF file and/or the Chain(s) of Custody. The RSCC will assign case numbers for non-ESAT projects and notify the Sample Custodian.

- 3.2.1.2 Log on to Labworks by double clicking on the Labworks icon on the Windows desktop. At the Labworks login prompt, enter in your name and password (already assigned by Region 9 Laboratory's Computer Specialist).
- 3.2.1.3 To enter a new case number into the database, single click on " Maintenance" from the Labworks Enterprise Desktop. From the drop down menu single click on "Case Number List". From the "Labworks Case Number Maintenance" screen, single click on the "Create/Modify Case Number" bar. This brings up the "Case Number Information Editing" options. In the "Header Info" screen, enter in the assigned case number and highlight the submittal date button. Click on the "User Fields" tab on the bottom of the screen and enter in the Site ID in the " Field Name" section. Next single click on the "Analysis Order" tab at the bottom of the page. If you know the analysis code, enter it in the field titled "Anl Code". If the analysis code is unknown, click on the first line of the "Anl Code" section. The next screen that appears is a scrolling list of all analysis codes and their descriptions. Scroll down the list until the appropriate analysis code is found. When it is highlighted, click on "OK", and repeat this process until all desired analysis codes are entered. Click on the "Save" button and then click on the "Exit" button.

3.2.2 Sample Information Entry/Sample Login

The following procedures include entering sample information into the Labworks database, assignment of laboratory identification numbers, printing of the Login Routing Sheet (Sample Login Report, Appendix F) and sample labels.

- 3.2.2.1 Log onto Labworks by double clicking on the Labworks Desktop icon. Enter your assigned user identification and password where indicated.
- 3.2.2.2 From the Labworks Desktop menu single click on "Login" and then single click on "Multi Sample Login" from the drop-down menu.

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- 3.2.2.3 Enter the following information on the Multi-Sample Login template:
 - -the case number
 - -the EPA ID (sample ID)
 - -the sample matrix (H2O, soil, sediment, etc.)
 - -Submit date
 - -collection date
 - -Sample Delivery Group (see section5.2.2.4.1)
 - -number of containers
 - -comments (if applicable)

This information must be entered for each sample so a unique laboratory tracking number can be assigned to each one. Make an entry for one additional sample (Lab Blank) per matrix, per SDG, (i.e. if 10 samples are received, 11 entries will be made).

- 3.2.2.3.1 SDGs are comprised of 20 samples from a given case (i.e., if 60 samples are received, there will be 3 SDGs). The SDG number is the day number of the year, preceded by the last 2 digits of the year, found on the "Activity Schedule, Optional Form 67" calendar. For example, June 5, 1998 is day 156. Therefore an SDG begun on this day would be 98156. If more than one SDGs are issued on a particular day, the SDG is to be further distinguished by a letter of the alphabet (98156b, 98156c, etc.).
- 3.2.2.3.2 If there is a 14 day gap from the last day a group of samples was received to the next delivery of samples from that case, then a new SDG is assigned.
- 3.2.2.4 Single click on the "Login" button on the bottom of the page. Copy the "Login Record File" number that appears in the box, on to the top of the Chain(s) of Custody.
- 3.2.2.5 A spreadsheet insert will appear on the screen showing the samples that were logged in and the corresponding Labworks issued ID numbers. Single click on the "OK" button. Next, single click on "File" on the menu on the top of the screen. Single click "Exit" on the drop-down menu.
- 3.2.2.6 From the Labworks Desktop main menu single click on "Maintenance". From the drop down menu single click on "Modify/Delete Sample".

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- 3.2.2.7 Select "Login Record File" from the "Sample Selection Options". Double click on the desired login record file and single click on the "OK" button.
- 3.2.2.8 To print sample labels, single click on the "Labels" button. A scroll down spreadsheet will appear displaying 6 columns. In the column titled "Count" type in the number of labels needed for each sample. Next, single click on the "OK" button. When asked if you want to select a printer, click on the "YES" button. In the "Select a Printer" box, highlight the *Epson FX-850* bar and single click on the "OK" button.
 - 3.2.2.8.1 Place the sample labels on the corresponding sample containers, being certain not to cover the EPA (field) IDs. Match the sample numbers on the Chain(s) of Custody with the numbers printed on the sample container. Double check each container after labels have been affixed to insure the right labels are on the right samples.
- 3.2.2.9 To print the Sample Login Report/Routing Sheet, double click on the Access Reports for Labworks button in the Labworks Enterprise group on the desktop screen. Single click on the Login Record File needed and single click on the Build the Sample Login Report button. Print the report.
 - 3.2.2.9.1 The Routing Sheet will print the case number, the SDG, the login date, the samples' Client ID and corresponding unique Laboratory ID, requested analysis, due date and matrix.
 - 3.2.2.9.2 When applicable, in handwritting, note that the pH was measured and make special note of any incorrect pH.
 - 3.2.2.9.3 When applicable, in handwritting, note any problems with samples, (i.e. incorrect labeling, broken containers, inefficient sample quantity, compromised sample integrity, etc.). Make this notation in the area designated as "comments" at the top of page 1.
- 3.3 Sample Login Report/Routing Sheet Distribution

Laboratory Routing Sheets, detailing the work for each laboratory department generated by Labworks are routed to the EPA and ESAT personnel responsible for the assigned

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analyses. The EPA Chemistry Team Leader receives a copy to check for login accuracy.

- 3.3.1 Group the Routing Sheets by analytical department (volatiles, metals, inorganic/general chemistry, etc.) With each Routing Sheet, attach a copy of the Chain (s) of Custody and the air bill.
- 3.3.2 Deliver one copy of the Laboratory Routing Sheet with attachments to the applicable analysts or technicians. Samples with short holding times (i.e., pH, nitrate) are a priority. Laboratory Routing Sheets and samples should be delivered to the analyst immediately.
- 3.3.3 Keep all original paperwork, including Routing Sheets, Chain(s) of Custody, air bills and any other applicable paperwork in the case logbooks stored in Room 503. There is a separate logbook for each EPA program (Superfund, RCRA, NPDES, WQM, etc.). Each logbook is kept in order by case numbers and date.
- 3.4 Sample Storage and Sign Out
 - 3.4.1 Soil and water samples are stored in the refrigerators located in Room 503. Tissue samples are stored in freezers in Room 503 or in Room 205.
 - 3.4.1.1 After samples have been logged in and labeled, place them on a shelf in one of the walk in refrigerators in Room 503 or in one the freezers in Room 503 or Room 205.
 - 3.4.1.2 All samples placed in a refrigerator or freezer must be signed into the Internal Chain of Custody logbook (Appendix G) located on the login bench directly across from the walk in refrigerators. Enter the lab identification number, their initials, the date the sample was logged into the storage unit, the refrigerator letter and the shelf number the sample(s) was placed.
 - 3.4.1.3 VOA samples are delivered to Room 201 by the Sample Custodian where they are stored and analyzed. VOA samples have a unique Internal Chain of Custody. Air samples are delivered to Room 203 where they are stored and analyzed. Air samples have a unique Internal Chain of Custody. In both cases, log samples into the Internal Chain of Custody at the time the samples are delivered.

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- 3.4.1.4 Most tissue samples are stored in Freezer Unit 21 in Room 503. Those samples are logged in and out of the regular Internal Chain of Custody logbook found on the login bench across from the cold rooms in Room 503. When Freezer Unit 21 reaches capacity, tissue samples are stored in Freezer Unit 3 in Room 205. Log samples stored in Room 205 into the Internal Chain of Custody logbook located on the bench to the right of Freezer Unit 3.
- 3.4.1.5 The refrigerators and freezer temperatures are monitored daily to insure they are operating correctly. This is performed by an ESAT employee assigned to the task of recording the temperature of each refrigeration unit in logbooks located in Room 503. Any observable irregularities are to be reported immediately to the building engineer.
 - 3.4.1.5.1 If a cold room or a freezer fails to maintain the samples at the required temperature, move the samples to another appropriate location (cold room/freezer), remembering to note the new location in the Internal Chain of Custody logbook.
- 3.4.2 To insure proper chain of custody procedure, samples must be signed out before being removed from Room 503 or Room 205 by all analysts and technicians. Prior to removing samples for analysis, the analysts or Technician must sign-out on the Internal Chain of Custody logbook. Signing-out samples is to include the date and the analysts initials. The samples must be returned and signed back in by the end of each working day.
 - 3.4.2.1 Review the Internal Chain of Custody logbook weekly to monitor the sign in/sign out process.
- 4.0 Sample Login Health and Safety Warnings

Samples received at the lab are of unknown toxicity and/or preserved with acids or bases that can harm the skin and eyes, therefore personal protective equipment is to be worn.

4.1 Lab coats, gloves and safety glasses are to be worn at all times while logging in samples.

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- 4.2 Upon opening the cooler, if any odor is detected, the cooler is to be promptly moved to the fume hood for unpacking, inspection and login.
- 4.3 Samples received in paint cans are to be moved to the fume hood for unpacking, inspection and login.

5.0 Materials and Apparatus

- 5.1 KayDry Tabletop soaker paper.
- 5.2 10 m/L disposable beakers.
- 5.3 ColorpHast indicator strips pH 0-14.
- 5.4 1:1 nitric acid (Reagent grade concentrated HNO3 and DI water).
- 5.5 1:1 sulfuric acid (Reagent grade concentrated H2SO4 and DI water).
- 5.6 3 m/L transfer pipettes.
- 5.7 Hach 2100N Turbimeter (see SOP 401).
- 5.8 -20 degrees C to 70 degrees C digital thermometer.
- 5.9 Cole Parmer 39650 Infrared thermometer.

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KEY	WO	RDS
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Sample Tracking, Sample Tracking Database, Chain-of custody, Sample

APPROVALS

APPROVED BY:

Management

DATE: 7/26/9

APPROVED BY:

Tion L. Ross

DATE: 7/19/99

EHAP Senior Scientist

APPROVED BY:_

CANDA TWA PAUM EHAP Quality Assurance Officer DATE: 7/15/99

PREPARED BY: Andrea Hoffman, Johanna Walters

DATE: 745 CA 7/

Environmental Hazards Assessment Program (EHAP) organization and personnel such as management, senior scientist, quality assurance officer, project leader, etc. are defined and discussed in SOP ADMIN002

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STANDARD OPERATING PROCEDURE Sample Tracking Procedures

1.0 INTRODUCTION

1.1 Purpose

This Standard Operating Procedure (SOP) discusses sample check-in and check-out procedures; the recording of chemistry data; sample disposal procedures; and the Sample Tracking Database.

1.2 Definitions

- **1.2.1** Sample is any environmental substance collected and analyzed for chemical content, toxicity, soil texture analysis, etc.
- 1.2.2 Sample Tracking Database is a relational database designed in Microsoft Access to trace a sample from the time it is checked into the storage facility until the sample is submitted to a laboratory for analysis or disposed of after a study is completed.
- **1.2.3** Chain-of-custody is a record describing in detail all pertinent information specific to each sample, including dates and signatures of persons handling the sample.
- 1.2.4 Sample Custodians are personnel, under direction of the lab liaison, responsible for receiving samples from field staff, delivering samples to the laboratory, and tracking samples in the Sample Tracking Database.

2.0 SAMPLE TRACKING

2.1 Sample Tracking Codes

Sample tracking codes are abbreviations for fields in the database that refer to specific information about each sample. The study number in combination with the sample number is identified as the key field and all information specific to the sample is referenced by the following codes back to the key field.

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SAMPLE CODES:

P= Primary

R= Replicate

B= Backup

FB= Field Blank

* = Split

S= Spike

BG= Background

BM= Blank Matrix

A= Acidified

U= Unacidified

RB= Rinse Blank

STORAGE LOCATION CODES refer to the storage location of each sample and the storage facility.

F= Fresno

R= Refrigerator

SR10= Sacramento Refrigerator #10

S= Sacramento

F= Freezer A= Air Temp SF05= Sacramento Freezer #05 SF06= Sacramento Freezer #06

W= Warehouse L= Lab

I=Ice Chest

SF07= Sacramento freezer #07

D= Deep Freeze

FZ= Freezesafe

SAMPLE TYPE CODES refer to the sample matrix collected.

FRU= Fruit

DVEG= Dislodgeable Vegetation

TWG= Twigs

SOI= Soil

SSS= Stainless Steel Sheets

EXT= Extract

WAT= Water

STD= Standard

VEG= Vegetation

SUR= Surrogate
TUR= Turf

SED= Sediment

FILT= Filtrate KIM= Kimbie

SAN= Sand

TAN= Tank AIR= Air

TRP= Air Cassettes

BRA= Branch

SAMPLE CONTAINER CODES refer to the type of container each sample is placed in during storage.

QMSJ= Quart Mason Jar

PMSJ= Pint Mason Jar

PBAG= Plastic Bag

FOIL= Aluminum Sheets

CAS= Air Cassettes

1LAMBR= 1 Liter Amber Bottle HPMSJR= Half Pint Mason Jar

HIVJAR= Hi-Vol Jar

P500mL= Plastic Bottle (500 mL) 1LPC= 1 Liter Polycarb. Bottle

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1LPP= 1 Liter Polyprop. Container
XADT= XAD Tube (small)
Summa= Summa Canister
HIV= High Volume Air Sampler_
500mLPC= 500mL Polycarb. Container
250mLAMBR= 250mL Amber Bottle
500mLAMBR= 500mL Amber Bottle
500mLHDPP= 500mL High Density Polyprop.

VIAL= Small Standard Vial XAD4= Large XAD 4 Tube LOV= Low Volume Air Sampler

LABORATORY CODES refer to the specific laboratory each sample is shipped to for analysis.

QUAN= Quanterra Laboratory
ATL= Aquatic Toxicology Lab
CDFG= CA Dept. of Food & Agr.
CDFG= CA Dept. of Fish & Game
CDFG= CA Dept. of Fish & Game
ALTA= ALTA Analytical Laboratory
VAL= Valent Dublin Laboratory
APPL= Apple Labs
MOR= Mores Laboratories Inc.
NCL= North Coast Labs
UCD= University California Davis
FRES= Fresno Soils Lab
WSAC= W. Sacramento Soils Lab

ANALYSIS TYPE refers to the type of test method to be performed on each sample.

C= Chemical F= Tracer E= Elisa
O= Organic P= pH M= Moisture
T= Texture B= Bulk Density V= Various

CHEMICAL ANALYSIS refers to the chemical analysis to be performed on each sample, if applicable.

OP=Organophosphate Screen

CB= Carbamate Screen

DI= Diazinon

EN/DI= Endosulfan/ Diazinon Screen

HEX=Hexazinone

TRI= Triclopyr

GLY= Glyphosate

TRIAZ= Triazine Screen

TOX= Biotoxicity

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TDM= Triclopyr, 2,4-D, MCPA

PIC= Chloropicrin MOL= Molinate

CARBO= Carbofuran

MeBr= Methyl Bromide

PROP= Propanil THIO= Thiobencarb

MP/MN= Methyl Parathion/Malathion

COMMENTS refers to any additional information regarding samples.

BS= Blind Spike

ACT TOX= Acute Tox

BB=Buck Brush BF= Bracken Fern EB= Elderberry DG= Deergrass

CHN TOX= Chronic Tox MB= Manzanita Berry

RD= Redbud

RB= Rinse Blank GF= Golden Fleece SR= Soap Root DB= Deer Brush PE= Pearly Everlasting

2.2 Sample Check-in Procedures

All samples received at the storage facility are immediately put in a refrigerator or freezer depending on the matrix specific storage requirements. The field crew fills out a three part check-in sheet (Figure A) using the sample tracking codes (Section 2.1).

The check-in sheet must be complete in order to properly track environmental samples. The following is a description of each key component of the check-in sheet.

Portion Filled Out By Field Staff

Project ID: The study number or name.

Date Received: The date the sample was received from the field crew. Checked-in by: The initials of the person who fills out the check-in sheet. Remarks: List ice chest number where samples were stored, Hobo Temp® temperature logger number (if necessary), and any additional or necessary information regarding the samples listed on the check-in sheet. For GLP studies, the ice chest number along with the maximum temperature samples were stored at in the ice chest must be marked on Hobo Temp® print-out as noted in SOP EQOT001.01. If temperature exceeded 6° C for refrigerated samples or 0° C for frozen samples, this must be documented on the sample check-in sheet in the comments section.

EHAP Sample No.: The number assigned to a labeled sample container.

Sample Code: List sample code (Section 2.1 for codes).

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Date Sample Collected: Note the sample collection date from the Chain-of-Custody.

Sample Type: Specify the type of sample collected (Section 2.1).

Container Type: What the sample is stored in (Section 2.1).

Analysis Type: The type of analysis the sample is intended for (Section 2.1).

Analysis: List the type of chemical or screen the sample is to be analyzed for.

Comment: Space provided for additional information regarding individual

samples (Section 2.1).

Portion Filled Out By Sample Custodian

Date/Logged in by: The date and person who enters information into the Sample Tracking Database.

Storage Location: List where the sample is being stored (Section 2.1).

After the check-in sheet is completed, the white and yellow copy are used to enter the information into the Sample Tracking Database and then filed with the QA/QC officer. The pink copy is given to the project leader in order to track ice chests and corresponding samples entering the storage facility (GLP studies only).

Each field sample is compared against it's corresponding Chain-of-custody (COC), then the COC is signed and dated by the person receiving the sample at the storage facility. The white and yellow copy of each COC is removed and sent with it's corresponding field sample to the laboratory. The pink COC copy is given to the Project Leader. Any remaining samples held at the storage facility are stored under their required storage conditions with the white and yellow copy of their corresponding COC's.

2.3 Sample Check-out Procedures

A three part check-out sheet is filled out for any sample leaving the storage facility (Figure B). The check-out sheet must be complete in order to properly track environmental samples leaving the storage facility. The check-out sheet is filled out by the sample custodian only.

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The check-out sheet is similar to the check-in sheet but differs in three components.

Date Delivered: The date the sample is taken to the laboratory.

Checked-out by: The initials of the person filling out and transporting the

sample to the laboratory.

Laboratory Delivering to: Specify the destination code for the sample

scheduled for analysis (Section 2.1).

A pink copy of the check-out sheet and the white and yellow copies of each COC are placed in a plastic bag and accompany samples transported to the laboratory. The samples are placed in ice chests and maintained at their required temperatures during transport using blue ice, wet ice or dry ice. The white and yellow copies of the check-out sheet are retained by the QA/QC officer and are used to enter information into the Sample Tracking Database.

2.4 Chemistry Results

After results are received from the laboratory, the laboratory sample number, and the extraction and analysis date for each sample are entered into the Sample Tracking Database using the appropriate Microsoft Access query.

2.5 Sample Disposal

After each study is completed, and with the approval of the Project Leader, all remaining samples stored in the storage facility may be disposed of by the sample custodian. A two part Sample Disposal Sheet is completed and includes information similar to the check-out sheet (Figure C). This information is then entered into the Sample Tracking Database using the appropriate Microsoft Access query. The white copy of the Sample Disposal Sheet is retained by the QA/QC officer while the yellow copy is used to enter the information into the database.

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3.0 Sample Tracking Database

All the information reported on the check-in, check-out, and sample disposal sheets is entered in the Sample Tracking Database using tables in Microsoft Access. Queries, forms and reports are designed specifically for each study to access fields for summarizing data.

3.1 Computer Generated Backups

Weekly backups are conducted by copying the database to a zip drive disk.

Study Number (Project ID):	Sample Tracking Staff Only:
Date Received (Warehouse):	Logged In By (data entry):
Checked-In By:	Data Entry Date:
Page of	Storage Location Code:

Remarks:

Samples were stored in ice chest #			at ch	eck-in.			
EHAP Sample #	Sample Code	Date Sample Collected	Sample Type	Container Type	Analysis Type	Analysis	Comments
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					'		
						·	

STATE OF CALIFORNIA

DEPARTMENT OF PESTICIDE REGULATION ENVIRONMENTAL HAZARDS ASSESSMENT PROGRAM

West Sacramento Field Office 3971 Commerce Drive, Suite D

SAMPLE CHECK-OUT SHEET

West Sacramento, CA 95691

(916) 322-3082

Study	y Number	(Project ID)):		Logged Out By (data entry):					
Date	Delivered	l:	-	ì	Data Entry Date:					
Chec	ked-Out I	Ву:		;	Storage Lo	cation Code:				
Labo	ratory De	livering To:]	Page of					
Remar	ks:									
EHAP Sample #	Sample Code	Date Sample Collected	Sample Type	Container Type	Analysis Type	Analysis	Comments			
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California Dept. of Pesticide Regulation Environmental Hazards Assessment Program 3971 Commerce Drive, Suite D West Sacramento, CA 95691 (916) 322-3082

Today's	Date	: <u> </u>
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Sample Disposal Sheet

Project ID	(Study no.	.):		Dis	posed by:		· 	
Date Dispos	sed:		Storage location:					
Remarks:	********	: # = # # # # # # # # # # # # # # # # #		2	== =======	********		
EHAP Sample #	Sample Code	EHAP Sample #	Sample Code	EHAP Sample #	Sample Code	EHAP Sample #	Sample Code	
								
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